



Herbicides in Lake Sediments: *Understanding Herbicide Fate*

Jason Ferrell, Mark Hoyer, Bill Haller

UF/IFAS Center for Aquatic and Invasive Plants

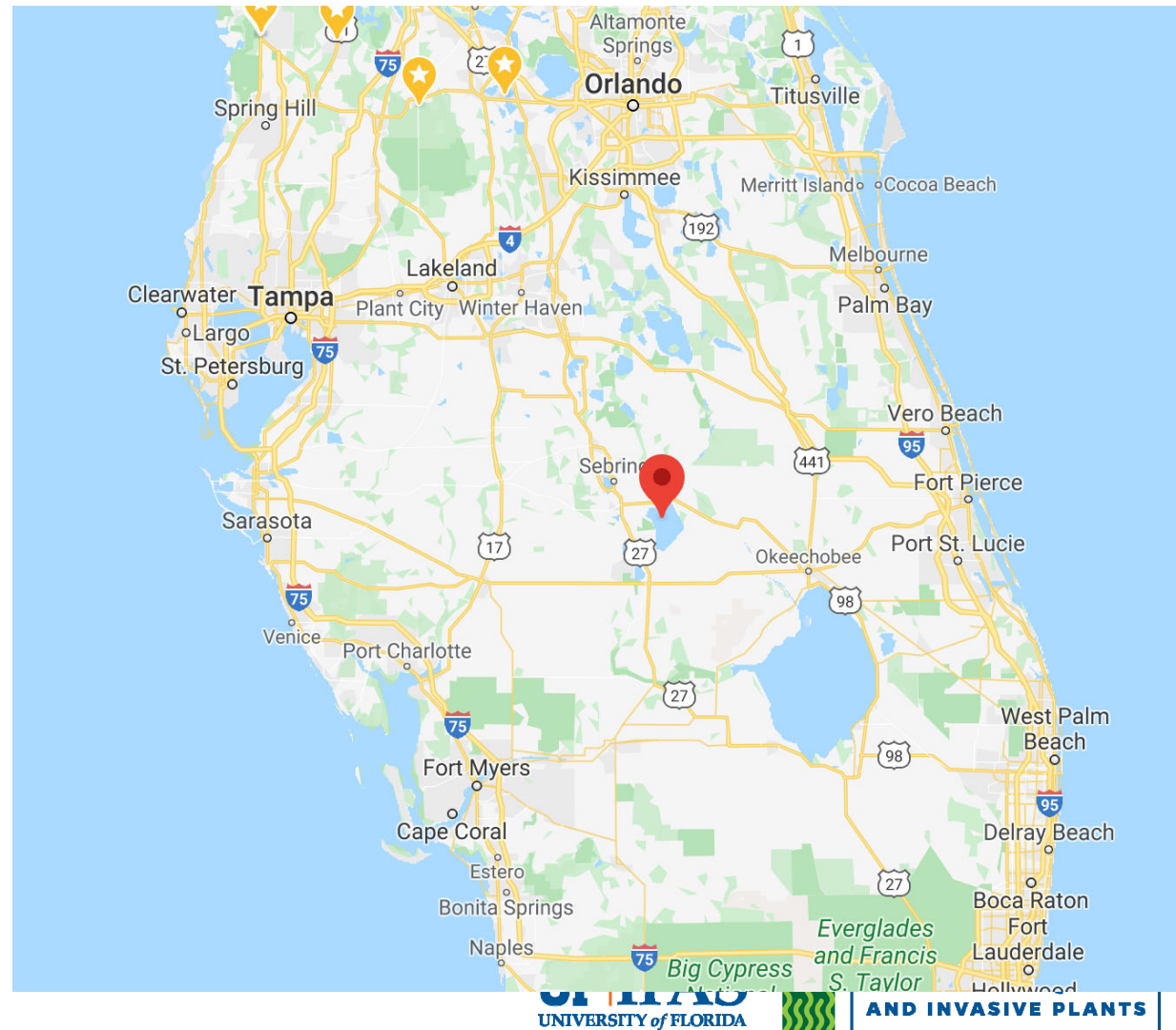


Legacy herbicides in lake sediments: *A Stakeholder Driven Approach*

Jason Ferrell, Mark Hoyer, Bill Haller – UF/IFAS

Lake Istokpoga, FL

- 27,000 acre lake in south FL
- Average depth: 4 ft
- Excellent fishery
- Historically infested with hydrilla

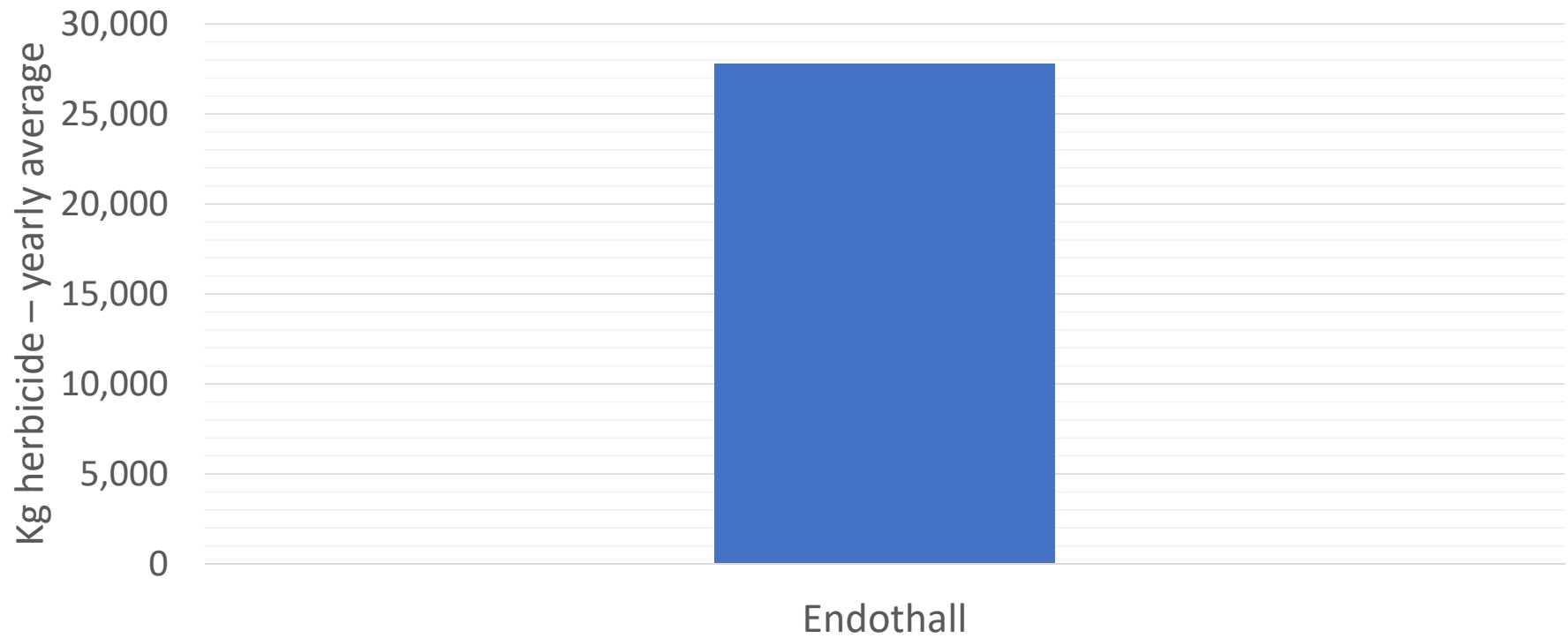


Hydrilla infestations

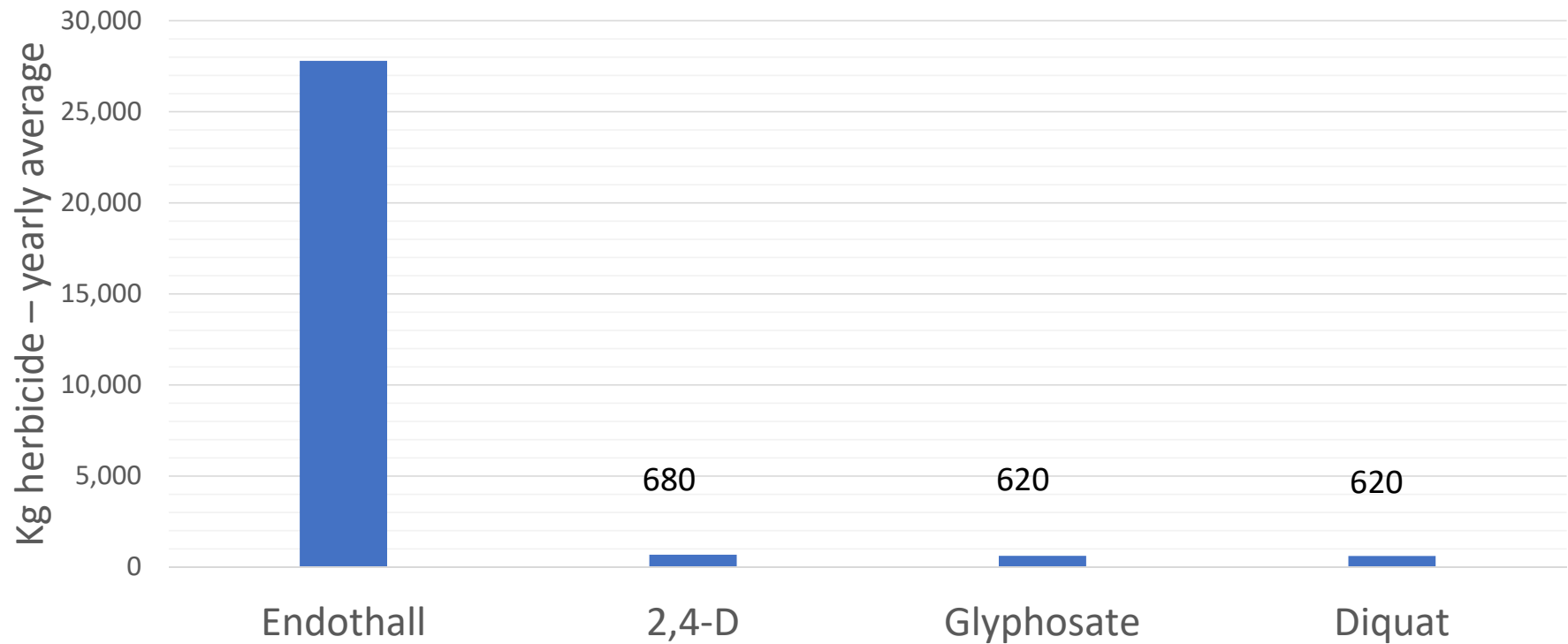
- In early 1990s, over 20,000 acres of hydrilla on the lake
- Aggressive, lake-wide, management instituted
- However, anglers have become convinced that hydrilla, is essential for a healthy fishery
- More recently, hydrilla is spot-treated



Herbicide use – 10-year average

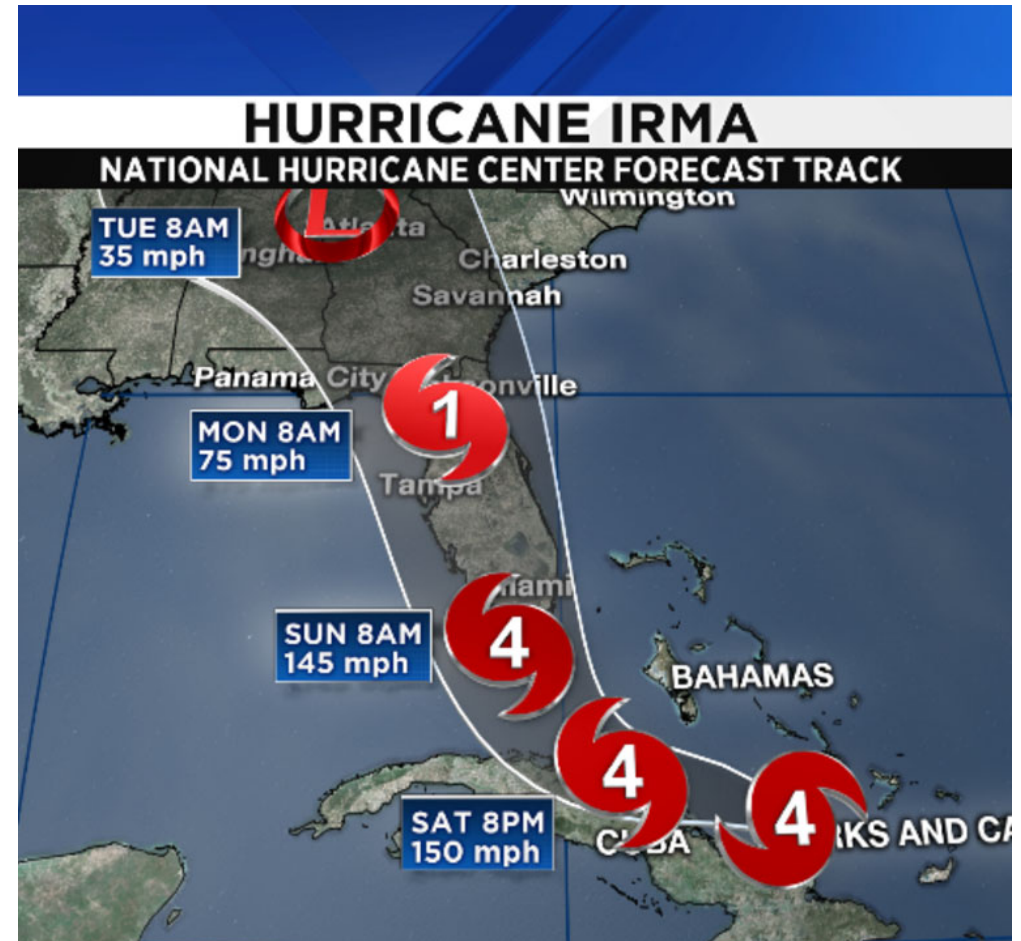


Herbicide Use – 10-year average



Hurricane Irma

- 2017 – massive disturbance
- Hydrilla was destroyed, didn't return
- Stakeholders believe contaminated sediment to blame for hydrilla failure to reestablish



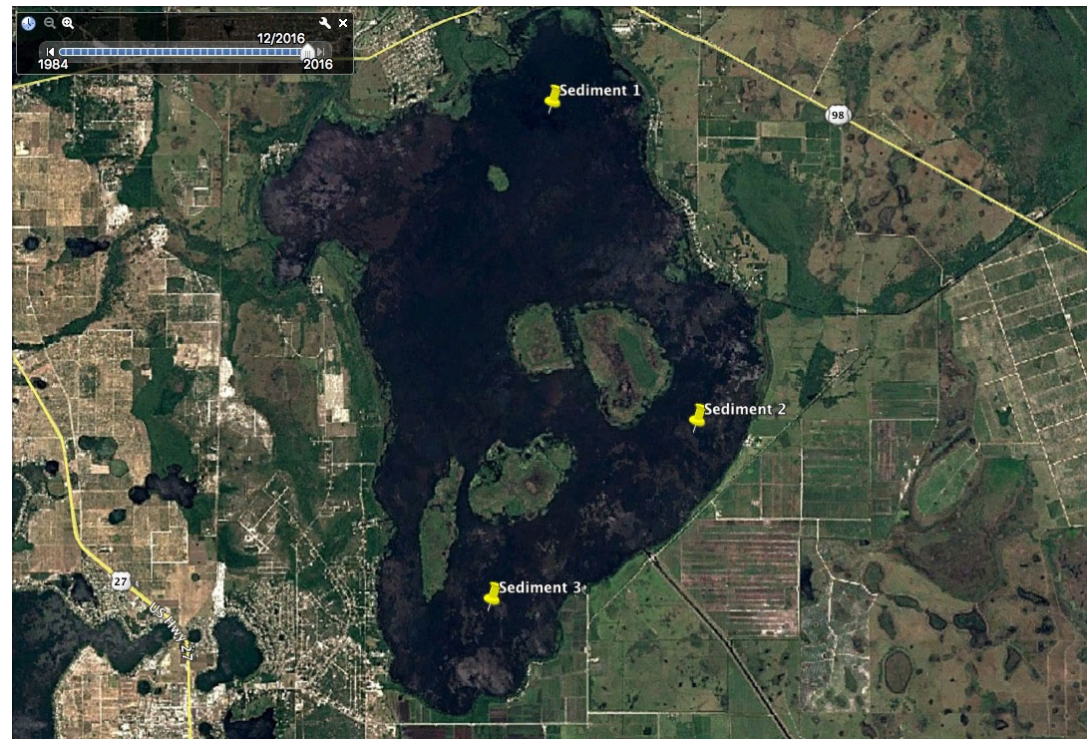
Lake Istokpoga Advisory Committee

- February 2019 – UF/IFAS Center for Aquatic and Invasive Plants became involved
- We recommended a 3-pronged approach
 - 1) Herbicide concentration in sediment study
 - 2) Bioassay sediment study
 - 3) Hydrilla tuber density study



Sampling locations

- We worked with the advisory committee to select 3 sites to sample at Istokpoga and Toho.



Sampling locations

- After the initial selection, the council wanted more sites
- We started with 36 locations, but we negotiated down to 9
- Some projects have just 3 sites and others have all 9



Lab analysis

- Samples taken at 9 spots
- Each sample was double bagged and stored on ice
- Shipped to a private lab
- Analyzed for 9 most commonly used herbicides on Lake Istok.



Bioassay

- Samples collected just as we did for the residue analysis
 - Members of advisory group were invited
- 5 gal of sediment from each station
- Sediment was de-watered and sifted



Bioassay

- The 9 Istokpoga samples were compared with 3 locations from Lake Toho
- We included 2 controls – pure sand and sand + potting mix.
- Tomato seeds and hydrilla tubers used
 - 4 replications used

Tuber sampling

- Original 3 locations were used
- 20 samples at each location
- Sampled every 30 m
- Samples sifted by hand and tubers identified by visual assessment



Results

Analytical analysis

Analytical analysis

Herbicide	Concentration*	Limit of detection (ppb)
Flumioxazin	U	20
Glyphosate	U	50
Imazamox	U	10
Imazapyr	U	10
Penxosulam	U	10
2,4-D	U	50
Triclopyr	U	50
Diquat	U	2000

*U = undetected

No lab could run endothall



Bioassay Sediment Results



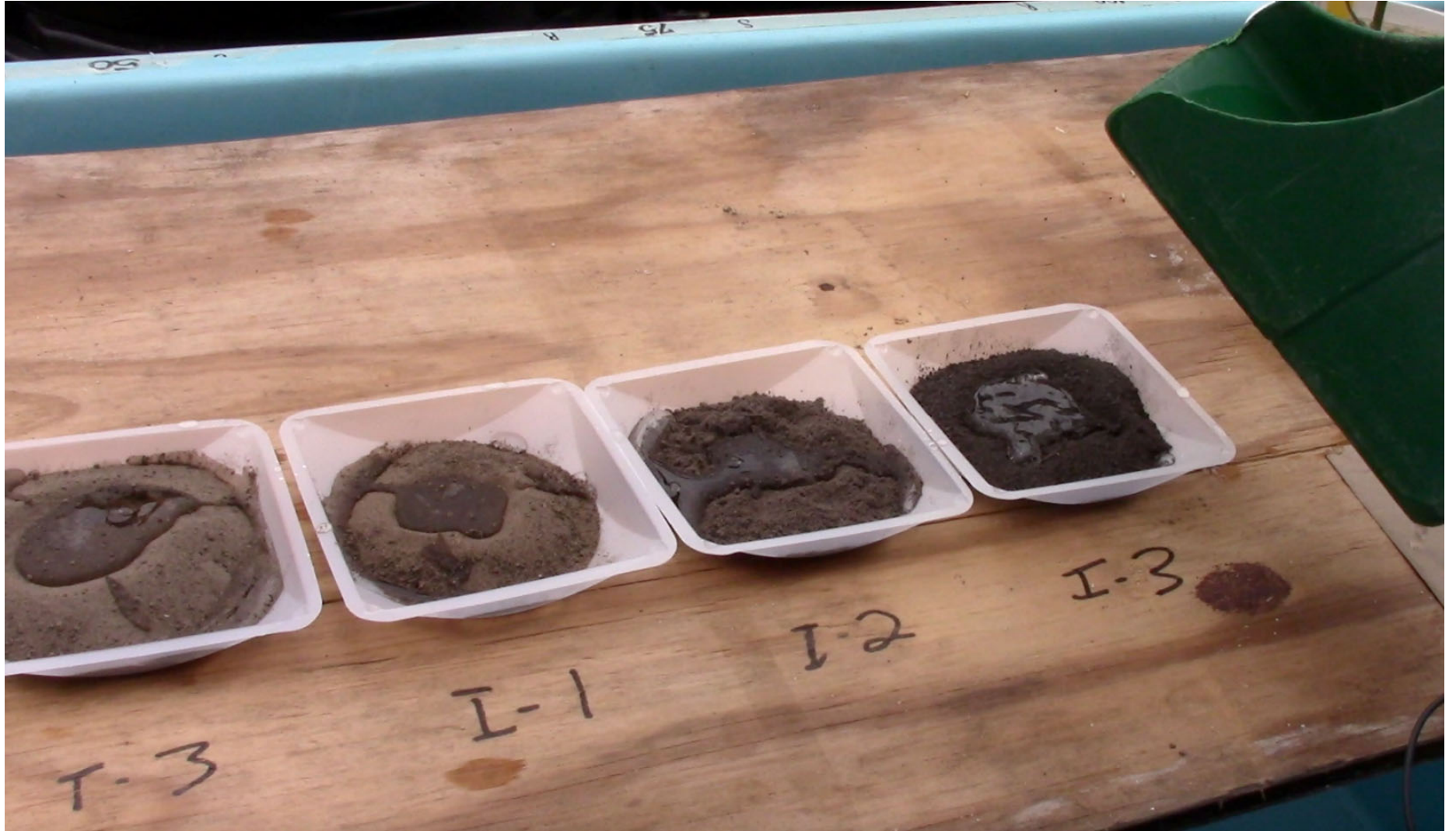
Bioassay results – Round 1

Location	Tomato germ (%)
Istok -1	72 ± 20
Istok -2	23 ± 5
Istok -3	88 ± 9
Toho -1	52 ± 17
Toho -2	70 ± 23
Toho -3	65 ± 23
Control (sand)	95 ± 5
Control (sand + OM)	92 ± 5

- Averages
- Control – 94%
- Istokpoga – 61%
- Toho - 63%

Bioassay results – Round 1

Location	Tomato germ (%)	Tomato wt (g)
Istok -1	72 ± 20	1.1 ± 0.1
Istok -2	23 ± 5	1.2 ± 0.1
Istok -3	88 ± 9	1.5 ± 0.2
Toho -1	52 ± 17	0.3 ± 0.1
Toho -2	70 ± 23	0.5 ± 0.1
Toho -3	65 ± 23	0.7 ± 0.1
Control (sand)	95 ± 5	0.9 ± 0.2
Control (sand + OM)	92 ± 5	



T-3

I-1

I-2

I-3

Bioassay results – Round 1

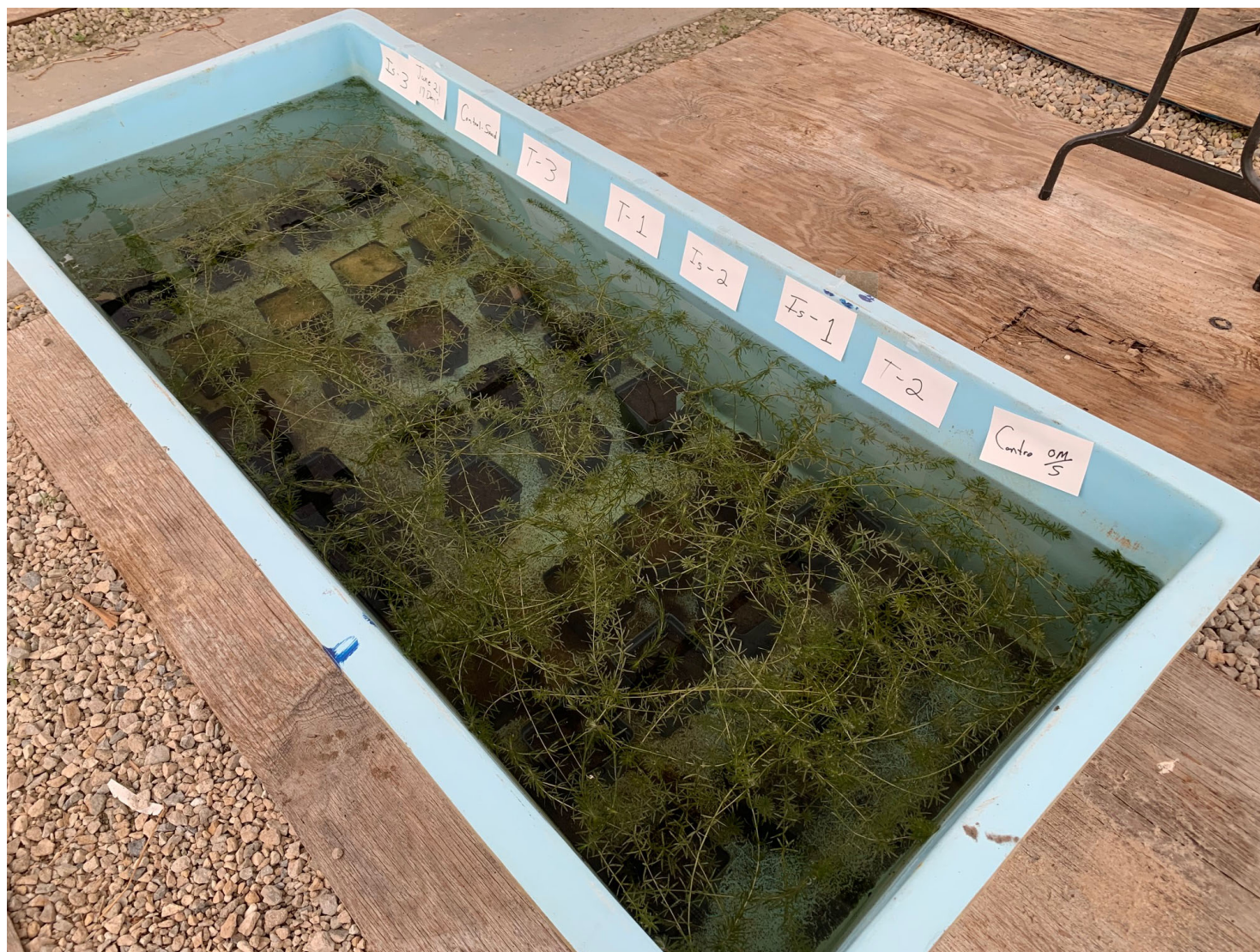
Location	Tomato germ (%)	Tomato wt (g)
Istok -1	72 ± 20	1.1 ± 0.1
Istok -2	23 ± 5	1.2 ± 0.1
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Toho -1	52 ± 17	0.3 ± 0.1
Toho -2	70 ± 23	0.5 ± 0.1
Toho -3	65 ± 23	0.7 ± 0.1
Control (sand)	95 ± 5	0.9 ± 0.2
Control (sand + OM)	92 ± 5	

Istokpoga: Bioassay Sediment Study



Bioassay results – Round 1

Location	Tomato germ (%)	Tomato wt (g)	Hydrilla germ (%)
Istok -1	72 ± 20	1.1 ± 0.1	79 ± 16
Istok -2	23 ± 5	1.2 ± 0.1	50 ± 13
Istok -3	88 ± 9	1.5 ± 0.2	48 ± 4
Toho -1	52 ± 17	0.3 ± 0.1	38 ± 20
Toho -2	70 ± 23	0.5 ± 0.1	38 ± 20
Toho -3	65 ± 23	0.7 ± 0.1	58 ± 16
Control (sand)	95 ± 5	0.9 ± 0.2	43 ± 8
Control (sand + OM)	92 ± 5		



Bioassay results – Round 2

Location	Tomato germ (%)	Tomato wt (g)	Hydrilla germ (%)
Istok -4	52 ± 22	-	-
Istok -5	82 ± 5	-	-
Istok -6	47 ± 32	-	-
Istok -7	40 ± 21	-	-
Istok -8	77 ± 15	-	-
Istok -9	27 ± 19	-	-
Control (sand + OM)	87 ± 9	-	-

Bioassay Sediment Study Conclusions

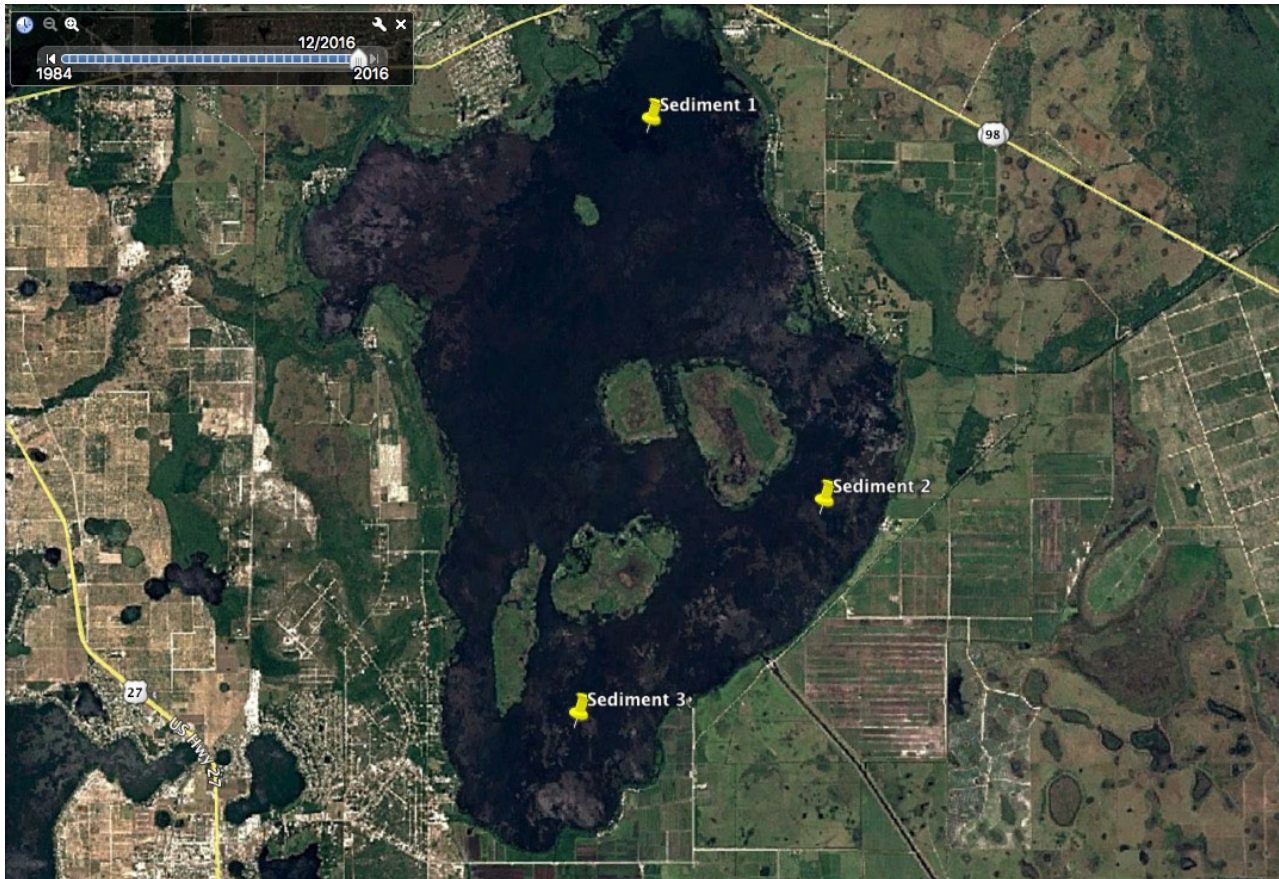
- Tomato seeds planted in Lake Toho and Lake Istokpoga sediments germinated equally to each other, but less than seeds in control soils
- Tomato dry weight was similar between Control (sand) and Istok 1,2,3; weight in Toho sediment was less.
- Hydrilla tubers planted in Lake Toho and Lake Istokpoga sediments grew equally to those planted in control soils

So what is going on?

- Lab can't detect herbicides in the sediments
- Tomato and hydrilla don't seem to detect herbicides
- Why is there no hydrilla in Lake Istokpoga?



Search for tubers in original 3 locations



Tuber numbers

- Location 1 – 0
- Location 2 – 0
- Location 3 – 0

- 60 cores – no tubers
- Istokpoga once had 20,000+ acres and 200-300 tubers per square meter is common. Where did they go?

Where did it go? We don't know

- Similar things have happened on Lake Weohyakapka
- 1995 – 6000A hydrilla on a 7500A lake
- 2004 – 3 hurricanes passed over, hydrilla disappeared and has not returned

Science vs social science

- I know herbicides don't accumulate in sediment. So, was this all a waste of time?
- The loudest voices at the beginning became the strongest allies.

Shout-out to Dr. Bill Haller



So, why don't these
herbicides build up?

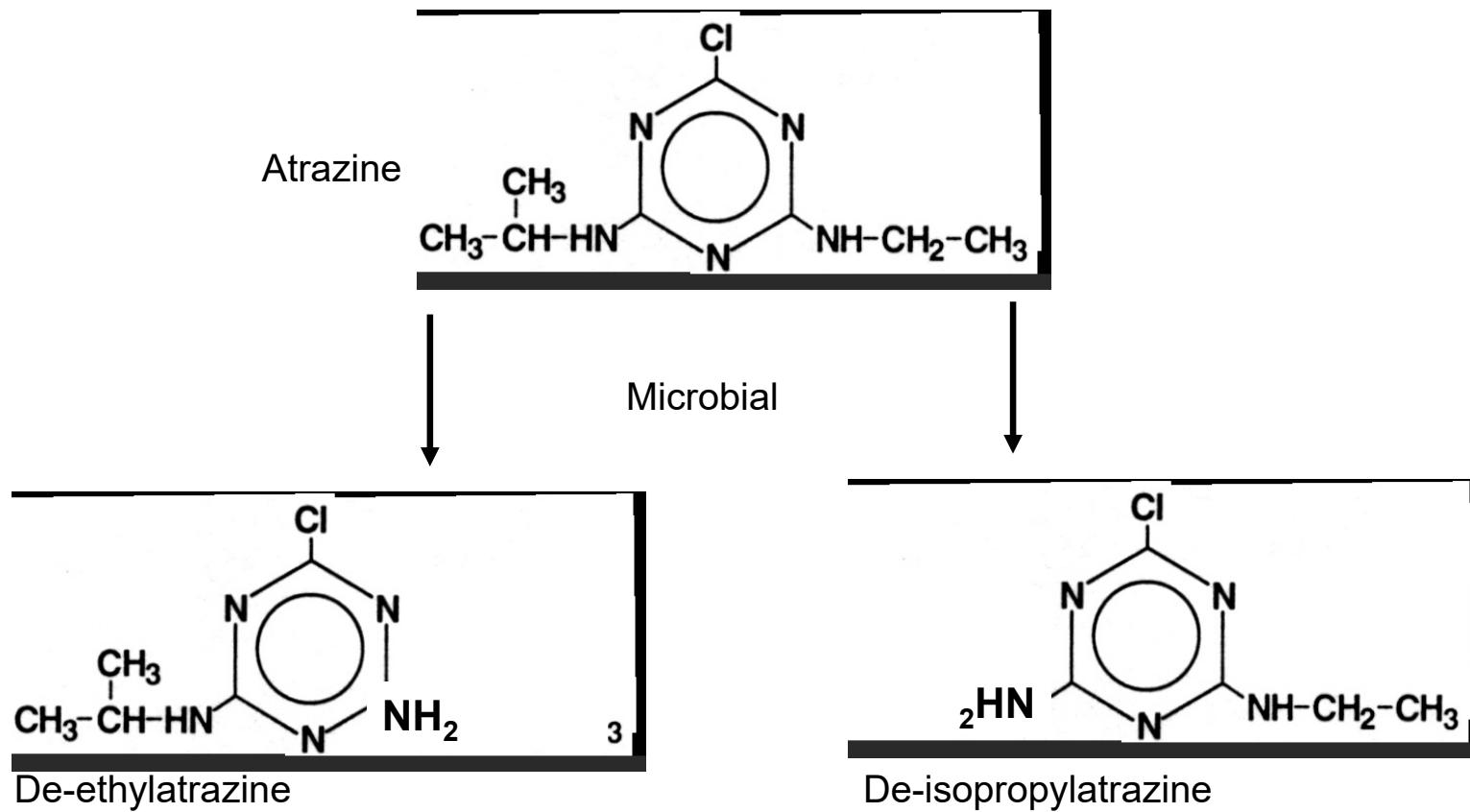
Herbicides in the Environment

What happens to them??

Herbicide Degradation

- How are herbicides degraded?
- Microbes – bacteria or fungi chew it up.
- Water – hydrolysis, water breaks it
- Light – photolysis, light breaks it

Atrazine: First step in degradation



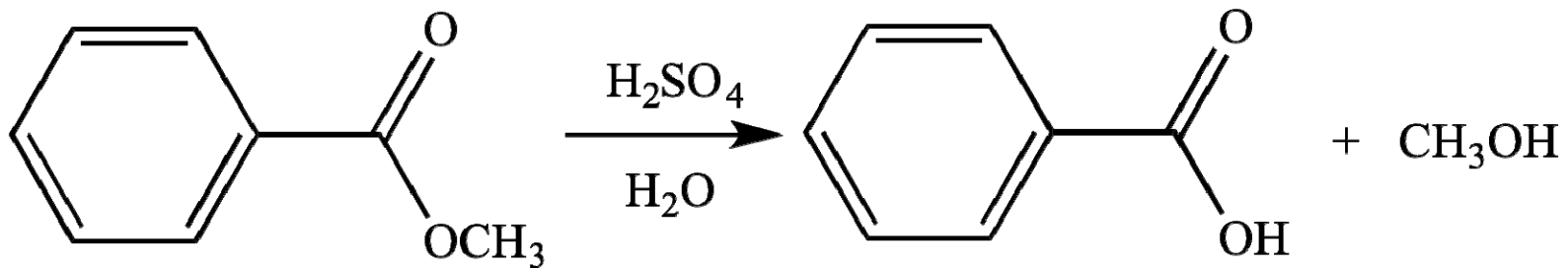
Microbial degradation

- Endothall
 - Works best in winter months
 - Breaks down to glutamic acid, an amino acid



Hydrolysis

- When water pH drives breakdown



- Particularly important for flumioxazin
 - Half-life: 16h at pH 7, 0.3h at pH 9

Photo-degradation

- Photolysis
 - Breakdown by sunlight

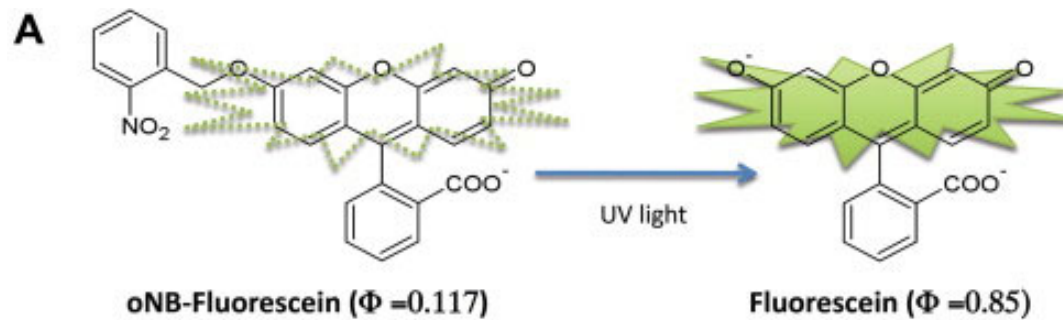
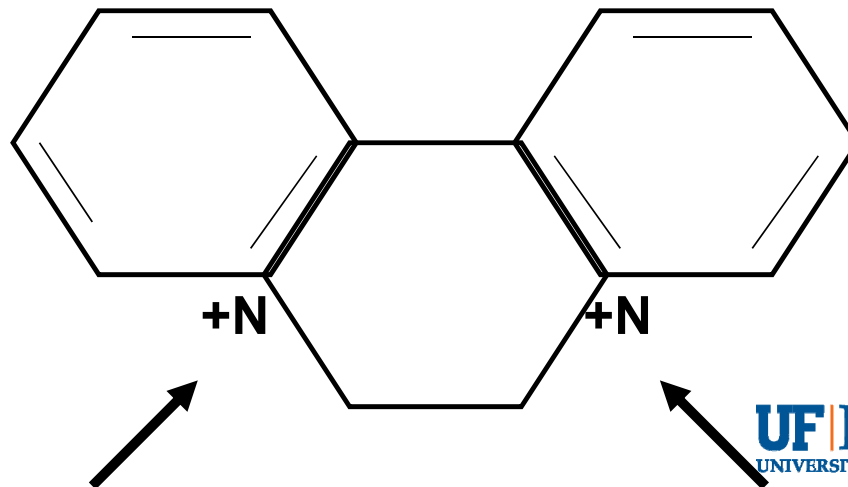


Photo-degradation

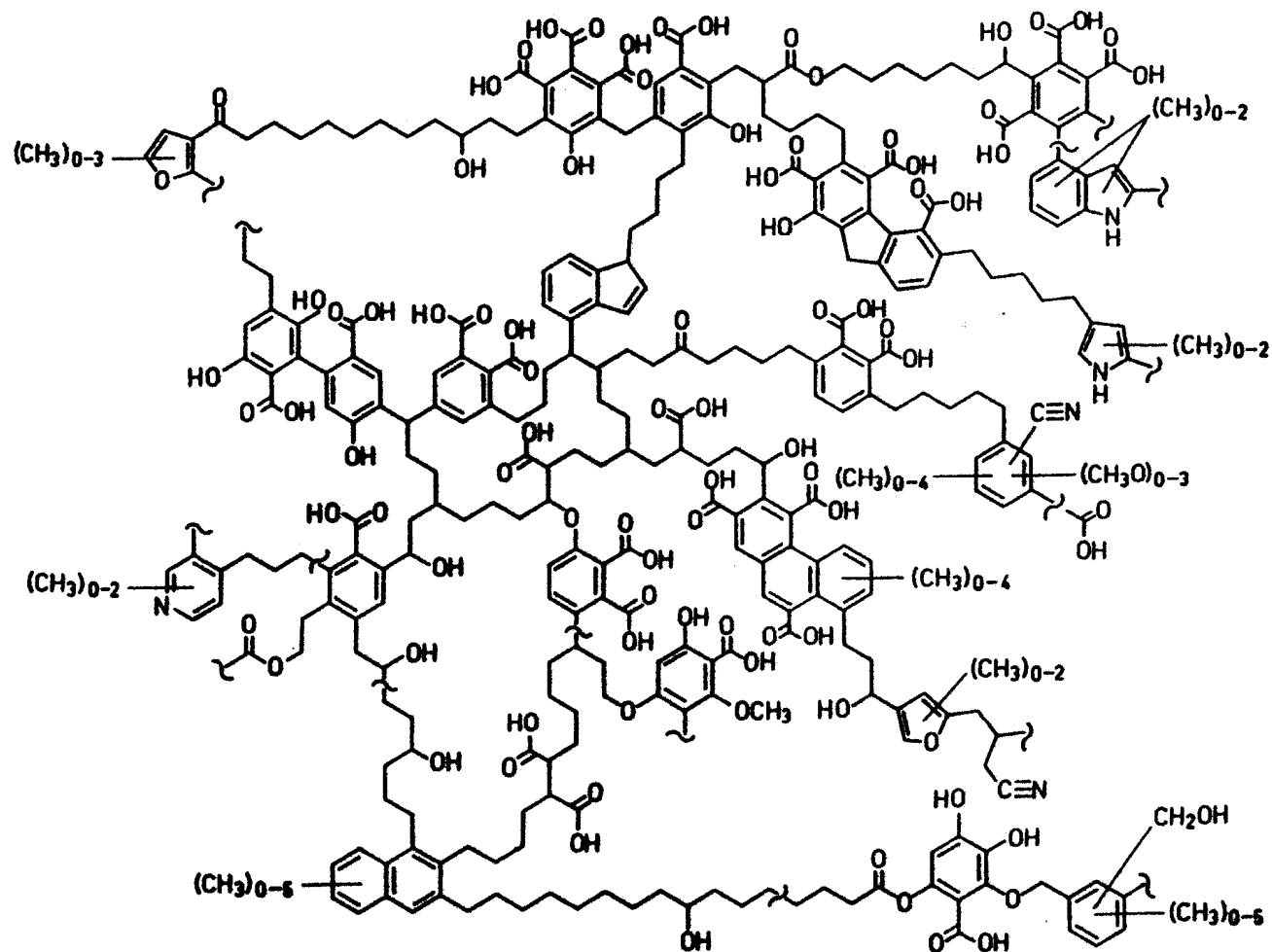
- Photolysis - Breakdown by sunlight
- Triclopyr
- Penoxsulam
- Imazamox
- Fluridone
- Diquat – to some degree

What Happens to Diquat?

- Breaks down from light and microbes
- Adsorption to soil or organic matter
 - Strongly sorbed, loves organic matter



Herbicide adsorption



Diquat

- When it binds to OM, it never lets go
- This protects the molecule from light and microbes
 - Microbes will eventually get it, but is very slow
- But...it also keeps it from being available to plants.

Testing for Diquat

- Very difficult process.
 - Only one lab in US does this.
 - Only guarantees 60% recovery.
- How does it work?
 - Take sediment and add highly concentrated sulfuric acid. Heat and stir. When OM is completely destroyed, you can find diquat.

But how do we know
they won't build up?

United States
Environmental Protection
Agency

Prevention, Pesticides
And Toxic Substances
(7101)

EPA 712-C-08-021
October 2008



Fate, Transport and Transformation Test Guidelines

**OPPTS 835.6200
Aquatic (Sediment)
Field Dissipation**

United States
Environmental Protection
Agency

Prevention, Pesticides
And Toxic Substances
(7101)

EPA 712-C-08-018
EPA 712-C-08-019
October 2008



Fate, Transport and Transformation Test Guidelines

**OPPTS 835.4300
Aerobic Aquatic
Metabolism**

**OPPTS 835.4400
Anaerobic Aquatic
Metabolism**